Iatrogenic phlebotomy loss resulting from the intensive clinical monitoring in the weeks immediately following birth remains the primary cause of neonatal anemia and the need for red-blood-cell (RBC) transfusion.

Reducing RBC transfusion needs in the neonatal intensive care unit (NICU) requires new approaches for reducing laboratory blood loss.

Fortunately, advances in laboratory technology involving benchtop analyzers, point-of-care testing devices, transcutaneous measurement and, to a lesser extent, the use of alternative specimens allow a greater number of analytes to be measured using increasingly smaller volumes of blood.

Indeed, these developments – along with more restrictive neonatal transfusion criteria – are responsible for the recent reductions observed in neonatal RBC transfusions in the NICU.

In the near future, more advanced technologies will offer even greater promise for preventing anemia, and thereby reduce the need for RBC transfusion in the NICU.

Anemic patients in the NICU are among the most highly transfused patient groups

Critically ill newborn infants are among the most heavily transfused groups [1]. Despite advances in the understanding of the pathophysiology and treatment of neonatal anemia, RBC transfusion continues to be the primary treatment for this commonly encountered condition.

Reducing blood transfusions is desirable, since they increase the risk of iatrogenic infection and adverse reactions. Although the use of increasingly restrictive transfusion criteria for neonates has been successful in reducing RBC transfusions [1, 2], still an estimated 1,000,000 transfusions are administered annually to neonates in the US [2, 3].
Approximately 60-80% of very-low-birth-weight (VLBW) infants (i.e., with birth weights <1,500 g) currently receive one or more RBC transfusions prior to hospital discharge as treatment for clinically significant anemia [4].

Although less well recognized, RBC transfusion of larger infants is also common, accounting for up to 45% of all transfusions in the NICU [5].

Laboratory blood loss in NICU: the primary cause of anemia and the need for RBC transfusions

There is strong agreement that the primary cause of neonatal anemia is iatrogenic phlebotomy loss resulting from the intensive monitoring which critically ill infants receive in the weeks immediately following birth [2, 3, 6].

Sampling loss in the NICU is directly associated with low gestational age and severity of illness [7]. Studies have shown that daily phlebotomy blood loss of 4-5% of an infant's 80 mL/kg blood volume during the early neonatal period is not uncommon among the sickest infants [8, 9].

When examined on a weekly basis for only premature infants, laboratory blood loss averages 15-30% of total blood volume every week for the first four weeks of life [3].

The volume of blood transfused in the NICU has been directly related to the volume of blood removed – in some cases on a 1-to-1, milliliter-for-milliliter basis [8-10].

In many [4] but not all [11] NICUs, approximately 50% of all RBC transfusions administered to VLBW infants are given in the first two weeks of life, with 70% administered by the first month [4], although iatrogenic blood loss continues to contribute to transfusion requirements beyond this early neonatal period.

Advances in laboratory blood testing have reduced anemia and transfusion in the NICU

Clearly, to reduce RBC transfusion in the NICU, new approaches to reducing iatrogenic blood loss for laboratory testing must be considered. It has been suggested that iatrogenic blood loss in neonates can be reduced by restricting blood testing to only that which is most essential [1, 2, 6].

This seemingly common-sense approach to preventing neonatal anemia is complicated by the fact that there is no consensus as to what constitutes “essential” testing. Moreover, there are no experimental data to show that this practice is either effective or safe.

In contrast, the technical improvements in the benchtop “analyzers” (instruments requiring ever-diminishing blood volumes) and the highly accurate bedside, point-of-care “monitors” (instruments that return the analyzed blood to the patient) described below have led to significant decreases in neonatal blood loss [12, 13].

Indeed, improvements in instrumentation in combination with the application of more restrictive RBC transfusion criteria are the primary reasons for the reduction in RBC transfusions reported for VLBW infants [1, 2].

Specific blood tests commonly ordered in the NICU for critically ill infants

What do we know about the kinds and frequency of blood testing in the NICU? This information would be helpful in designing effective strategies for reducing iatrogenic neonatal blood loss.

For all critically ill patients – including neonates – repeated measurement of blood gases, electrolytes, glucose and hemoglobin concentration from indwelling arterial or central venous vascular catheters remains a cornerstone of the intensive care that such individuals require.

To determine the number and kinds of laboratory blood tests typically performed on critically ill VLBW infants with indwelling umbilical arterial catheters in the first weeks of life, we retrospectively examined the specific repetitive laboratory tests that a group of 50 VLBW infants experienced [14].
As illustrated in Figure 1, the number of laboratory blood tests performed during the first week of life while arterial catheters were available for blood sampling averaged ~20 per infant per day.

The most commonly performed blood tests in our NICU were blood gases and electrolytes, followed in decreasing order by glucose, hemoglobin/hematocrit, total bilirubin and calcium.

Hence, reducing iatrogenic neonatal loss should primarily focus on reducing the blood required for performing these tests.

Expanding the capabilities of these instruments by including additional tests such as electrolytes, lactate, bilirubin, etc. has resulted in little-to-no increase in sample volume [16].

2. Point-of-care-testing (POCT) analyzers and monitors

Like benchtop blood gas analyzers in the central core laboratory, modern POC devices including “analyzers” (which permanently remove blood) and in-line “monitors” (which return the analyzed blood to the patient) are also capable of analyzing whole-blood samples in small quantities.

Since with POC devices all steps in the processing and handling (including specimen labeling) of samples can be performed at the bedside by a single individual, an important feature of the POC devices is that preanalytical error is less than for core laboratory analyzers [17].

In addition to conserving specimen, POC testing permits rapid decision making as a result of immediate access to test results.

The menus available for POC analyzers – but not yet for monitors – are substantial and growing, and include many of the most commonly ordered tests included in the Figure.
lab, and the burden of POC testing often falls on the nursing staff, who are already coping with multiple, complex patient care duties and who require formal training in the use of the equipment.

However, quality test results are achievable through oversight and partnership with the laboratory, and the increased cost may be more than offset by the prospect of better patient outcomes.

### 3. Transcutaneous measuring devices

Transcutaneous measurements will decrease the need for blood samples. The thin, translucent skin of newborns makes the measurement of some analytes easier than in adults [18]. For example, measurement of $pO_2$ and $pCO_2$ are available transcutaneously and have been shown to correlate with assays performed on a blood gas analyzer [19].

Despite this advantage, the use of transcutaneous measurements has declined following the introduction of pulse oximetry $SpO_2$ measurement. Although pulse oximetry is inaccurate at high $pO_2$ levels, it is more user-friendly for the nurse and technician and it does not carry the risks of thermal injury.

### Table 1: Technologies for minimizing laboratory blood loss in NICU environment

<table>
<thead>
<tr>
<th>Laboratory method</th>
<th>Range of blood volumes</th>
<th>Analytes</th>
<th>Advantages</th>
<th>Disadvantages</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>POC &amp; whole-blood benchtop analyzers</td>
<td>25-100</td>
<td>Na, K, Hb/Hct, blood gases &amp; pH, glucose, bilirubin &amp; most others</td>
<td>Small whole-blood samples; Expanding menu; Rapid analysis; Less preanalytical error in patient identification (POCs only)</td>
<td>For some POCs: 1) connectivity to computer systems to patient's medical record; and 2) oversight and regulations for CLIA non-waived testing (in the US)</td>
<td>[17, 25]</td>
</tr>
<tr>
<td>In-line POC</td>
<td>0-24</td>
<td>Na, K, Hb/Hct, blood gases &amp; pH</td>
<td>Little to no blood loss</td>
<td>Requires indwelling arterial catheter; Limited menu; Expensive; Connectivity to computer systems</td>
<td>[13, 26]</td>
</tr>
<tr>
<td>Transcutaneous</td>
<td>None</td>
<td>$SpO_2$, $tcpO_2$, $tcpCO_2$ &amp; bilirubin</td>
<td>Requires no blood sampling; Continuous for the gases</td>
<td>Limited menu</td>
<td>[18, 19]</td>
</tr>
<tr>
<td>Alternative body fluids (e.g., saliva, interstitial fluid)</td>
<td>10-100</td>
<td>Cortisol &amp; some drugs</td>
<td>Requires no blood sampling; Conserves blood</td>
<td>Limited reference values; Limited menu; Interstitial fluid requires indwelling capillary tube</td>
<td>[20]</td>
</tr>
<tr>
<td>Future technologies</td>
<td>None</td>
<td>Glucose, tissue Hb &amp; tissue oxyhemoglobin saturation</td>
<td>Utilize nano-technologies, allow continuous monitoring; and/or require no blood sampling</td>
<td>Not commercially available</td>
<td>[22, 24]</td>
</tr>
</tbody>
</table>
More recently, reliable transcutaneous measurement of bilirubin has become possible. Several meters are commercially available that have demonstrated good agreement of serum and transcutaneous bilirubin measurements.

As with all laboratory testing, proper attention to details such as calibration and ongoing comparisons with the serum method is essential.

Thus far, transcutaneous bilirubin measurement has been reserved primarily for screening purposes; transcutaneous bilirubin levels that require clinical intervention should still be confirmed on a blood sample using a chemical method.

4. Use of alternative specimens
Saliva and interstitial fluid are the two most studied – yet underutilized – alternative specimen types. Saliva has proven feasible and reliable for the measurement of hormones such as cortisol and some commonly administered neonatal drugs, e.g., theophylline and caffeine [20].

Specialized devices to collect both types of fluid are available [21].

Problems include sensitivity of assays used, since the concentration of some analytes in saliva may be far less than in blood. For this reason, saliva specimens must be free of blood. Reference ranges for saliva are less well established than those for blood, especially for neonates.

5. Future technologies
Non-invasive measurement of analytes in the circulation has long been a goal of researchers and clinicians. Infrared spectrophotometric methods for non-invasive glucose, tissue Hb and oxyhemoglobin saturation determinations have been described for some time [22, 23] and are likely to come on the market in the future.

In a few years, microchip, microarray and nanotechnology devices performing PCR, immunoassay and other analytical procedures on sub-microliter samples will also become available [24].

This technology will open up whole new areas for diagnosis and treatment of the neonate. Finally, a new POC blood counter capable of performing hemoglobin, hematocrit, WBC and three-point differential on 20 µL of capillary blood (as opposed to large hematology analyzers requiring 10 times this amount) is available in Europe and has been submitted to the FDA for approval.

In conclusion, advances in laboratory technology will continue the trend of allowing for the determination of more analytes on smaller and smaller volumes of blood.

Reducing the transfusion needs of critically ill neonates will improve their treatment by preventing the development of clinically significant anemia attributable to laboratory phlebotomy loss.
References